

PLANT SCIENCE

Make Your Plant Research Easy & Cost-Effective

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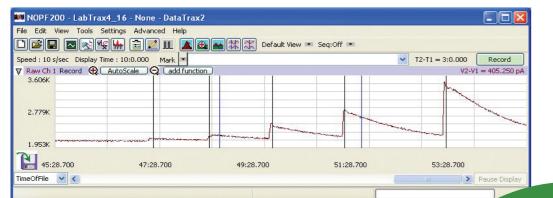
SOLUTIONS FOR THE STUDY OF PLANTS

FREE RADICAL ANALYSIS FOR PLANT STRESS RESEARCH

WPI Free Radical Analyzer (4-channel TBR4100 and single-channel TBR1025) and the LabTrax Data Acquisition System with a range of biosensors, enable real-time, highly sensitive detection of reactive oxygen species (ROS) including free radicals such as O_2 , NO, H_2O_2 , H_2S and CO in plants. Generation of reactive oxygen species (ROS) is one of the most common response to various stresses (biotic and abiotic) encountered by plants.¹

ROS are important physiological indicators of intracellular cell signaling and homeostasis. They are generated as byproducts of photosynthesis and respiration, which are localized in chloroplasts, mitochondria and peroxisomes. They play an important signaling role in controlling processes such as growth, development, response to biotic and abiotic environmental stimuli, and programmed cell death.² They also induce oxidative damages under several environmental stress conditions including salinity, drought, cold, heavy metals, UV irradiation, etc. when the balance between ROS production and elimination is disturbed.³ Plants have an innate capability to biosynthesize antioxidants to scavenge ROS. This makes plants a rich source of antioxidants with great therapeutic applications.⁴

WPI Free Radical Analyzers have been widely used for measurement of free radicals in various studies related to ROS. ⁵⁻¹¹



The output shows the raw data for an ISO-NOPF200 (NO sensor). Only one of the four channels is used in this application.



This is a typical laboratory setup of a WPI free radical analyzer with data acquisition system.



PLANT ELECTROPHYSIOLOGY

Plant electrophysiology is the study of the electrochemical phenomena associated with biological cells and tissues in plants. The conduction of bioelectrochemical excitation is a rapid method of long-distance signal transmission between plant tissues and organs. Plants promptly respond to the following changes:

- Luminous intensity
 Mechanical stimulation
- Osmotic pressure
 Plant growth stimulants
- Temperature
- Cutting
- Water availability
- Wounding
- in soil

Salts & mineral concentration

• Chemical compounds like

herbicides

Soil water concentration

Every response initiates an electrical impulse that is propagated to adjacent excitable cells. The bioelectrochemical system in plants not only regulates stress responses, but also photosynthetic processes. The generation of electrical gradients is a fundamental aspect of signal transduction. There are two major divisions of cellular electrophysiology, intracellular recording and extracellular recording.¹⁵

WPI amplifiers have been extensively used and cited for electrophysiological studies for both intracellular and extracellular recording. $^{\rm 16,\,17}$

HORTICULTURAL INSTRUMENTS

We offer a unique assortment of instruments to aid on plant physiology, plant molecular, cellular and developmental biology.

	15917	Iris Forceps, 10 cm, Curved, 1x2 Teeth
	15914	Iris Forceps, 10 cm, Straight, Serrated
	504473	Nugent Utility Forceps, 10 mm, Angled Tip
	500456	Filter Forceps, 11 cm, Straight
	500457	Filter Forceps, 11 cm, Bent Flat Jaw
	503411	Horticultural Forceps, 12.5 cm
	501241	Kelly Hemostatic Forceps, 14 cm, Straight
	501715	Kelly-Rankin Hemostatic Forceps, 15.5 cm, Curved
	501714	Kelly-Rankin Hemostatic Forceps, 15.5 cm, Straight
	501708	Rochester-Pean Hemostatic Forceps, 19 cm, Curved
	504639 504642	Reusable Rapid Punch Kit with 0.5 mm Tip Replacement Tip for Rapid Punch size 0.5 mm
	WPB315AS	Black Coated, 11 cm (4.3 in.), EZ Lever, Strong Blades, 6.5 mm from Tip to Bend
	WPB315AB	Black Coated, 11 cm (4.3 in.), EZ Lever, 4.5 mm Tip

OTHER INSTRUMENTS & ACCESSORIES

WPI offers a wide range of instruments that can be used for plant protection research and plant manipulation. WPI carries dissecting instruments or tools, inverted microscopes, syringe pumps that can inject in ranges from nanoliters to milliliters, micropipettes puller that can produce micropipettes with tip diameters in the range from sub-micron tips to 10um (using a beveler).



The WPI micropipette puller is a compact, versatile and reliable workhorse.



WPI offers Thin-wall or standard borosilicate glass in different outer diameter with or without filament.

BEST SELLER



MICROMANIPULATORS

• High precision

Manual

- Left or right-handed orientation
- Ranges from 3 mm travel to 37 mm travel depending on the axis (x, y and z)
- Variety of stands magnetic or tilt-base
- MOTORIZED
- 3-axis manipulators (x, y and d (diagonal))
- 4-axis manipulators (x, y, z and d)
- Ergonomic hand controller or software controlled
- Ranges from 25 mm on the x, y and z and 50 mm diagonally

REFERENCES

- Ines Ben Rejeb, Victoria Pastor and Brigitte Mauch-Mani. Plant Responses to Simultaneous Biotic and Abiotic Stress: Molecular Mechanisms. Plants 2014, 3, 458-475; doi:10.3390/ plants3040458
- 2 Julia Bailey-Serres, Ron Mittler. Plant Physiology, June 2006, Vol. 141, pp. 311, www.plantphysiol.org.
- 3 Kaushik Das and Aryadeep Roychoudhury. Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. Front. Environ. Sci., 02 December 2014 | https://doi.org/10.3389/fenvs.2014.00053
- 4 Deepak M. Kasote, Surendra S. Katyare, Mahabaleshwar V. Hegde, and Hanhong Bae. Int J Biol Sci. 2015; 11(8): 982–991.
- 5 Silveira, N. M., Seabra, A. B., Marcos, F. C. C., Pelegrino, M. T., Machado, E. C., & Ribeiro, R. V. (2019). Encapsulation of S-nitrosoglutathione into chitosan nanoparticles improves drought tolerance of sugarcane plants. Nitric Oxide, 84, 38–44. https://doi.org/10.1016/J. NIOX.2019.01.004
- 6 da Silva, C. J., Batista Fontes, E. P., & Modolo, L. V. (2017). Salinity-induced accumulation of endogenous H2S and NO is associated with modulation of the antioxidant and redox defense systems in Nicotiana tabacum L. cv. Havana. Plant Science, 256, 148–159. https:// doi.org/10.1016/j.plantsci.2016.12.011
- 7 Oliveira, H. C., Gomes, B. C. R., Pelegrino, M. T., & Seabra, A. B. (2016). Nitric oxide-releasing chitosan nanoparticles alleviate the effects of salt stress in maize plants. Nitric Oxide, 61, 10–19. https://doi.org/10.1016/j.niox.2016.09.010
- 8 Silveira, N. M., Frungillo, L., Marcos, F. C. C., Pelegrino, M. T., Miranda, M. T., Seabra, A. B., ... Ribeiro, R. V. (2016). Exogenous nitric oxide improves sugarcane growth and photosynthesis under water deficit. Planta, 244(1), 181–190. https://doi.org/10.1007/s00425-016-2501-y
- 9 Yarmolinsky, D., Brychkova, G., Kurmanbayeva, A., Bekturova, A., Ventura, Y., Khozin-Goldberg, I., ... Sagi, M. (2014). Impairment in Sulfite Reductase Leads to Early Leaf Senes-

cence in Tomato Plants. Plant Physiology, 165(4), 1505–1520. https://doi.org/10.1104/ pp.114.241356

10 Diniz, T., Pereira, A., Capettini, L., Santos, M., Nagem, T., Lemos, V., & Cortes, S. (2013). Mechanism of the Vasodilator Effect of Mono-oxygenated Xanthones: A Structure-Activity Relationship Study. Planta Medica, 79(16), 1495–1500. https://doi.org/10.1055/s-0033-1350803

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Magnetic stand

- 11 Pandolfi, C., Pottosin, I., Cuin, T., Mancuso, S., & Shabala, S. (2010). Specificity of Polyamine Effects on NaCl-induced Ion Flux Kinetics and Salt Stress Amelioration in Plants. Plant and Cell Physiology, 51(3), 422–434. https://doi.org/10.1093/pcp/pcq007
- 12 Narendra Tuteja & Shilpi Mahajan. Calcium Signaling Network in Plants. An Overview. Plant Signal Behav. 2007 Mar-Apr; 2(2): 79–85.
- 13 Pérez-Bueno ML, Pineda M, Cabeza FM and Barón M (2016)Multicolor Fluorescence Imaging as a Candidate for Disease Detection in Plant Phenotyping. Front. Plant Sci. 7:1790.doi: 10.3389/fpls.2016.01790
- 14 E.H. Murchie, T. Lawson. Chlorophyll fluorescence analysis: a guide to good practice and understanding some new applications. Journal of Experimental Botany, Volume 64, Issue 13, October 2013, Pages 3983–3998, https://doi.org/10.1093/jxb/ert208
- 15 Plant Electrophysiology Signaling & Responses. Alexander G Volkov, Editor. Springer Publication. ISBN 978-3-642-29110-4 (eBook)
- 16 Chatterjee SK, Das S, Maharatna K, Masi E, Santopolo L, Mancuso S, Vitaletti A. Exploring strategies for classification of external stimuli using statistical features of the plant electrical response. J.R. Soc. Interface 12: 20141225. (2015) http://dx.doi.org/10.1098/rsif.2014.1225
- 17 Jens B. Hafke, Sabina-Roxana Höll, Christina Kün and Aart J. E. van Bel. Electrophysiological approach to determine kinetic parameters of sucrose uptake by single sieve elements or phloem parenchyma cells in intact Vicia faba plants. Front. Plant Sci., 31 July 2013 | https:// doi.org/10.3389/fpls.2013.00274

WORLD PRECISION INSTRUMENTS

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